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INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

DRAFT

TIMOTHY

UPOV Code(s):

PHLEU_BER;
PHLEU_PRA*Phleum bertolonii* DC.;
Phleum pratense L.

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

*prepared by experts from Slovakia
to be considered by the
Technical Working Party for Agricultural Crops
at its forty-eighth session, to be held in Montevideo, Uruguay,
from 2019-09-16 to 2019-09-20*

Disclaimer: this document does not represent UPOV policies or guidance

Alternative names:*

Botanical name	English	French	German	Spanish
<i>Phleum bertolonii</i> DC.	Diploid Timothy, Small Timothy, Smaller Cat's-tail; Timothy; Turf Timothy	Fléole diploïde; Petite fléole	Zwiebellieschgras	Fleo
<i>Phleum pratense</i> L., <i>Phleum intermedium</i> Jord., <i>Phleum</i> <i>parnassicum</i> Boiss., nom. nud.	Meadow cat's-tail, Timothy	Fléole des prés	Timothe, Wiesenlieschgras	Fleo de los prados

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website (www.upov.int), for the latest information.]

TABLE OF CONTENTS	PAGE
1. SUBJECT OF THESE TEST GUIDELINES.....	4
2. MATERIAL REQUIRED.....	4
3. METHOD OF EXAMINATION.....	5
3.1 Number of Growing Cycles.....	5
3.2 Testing Place.....	5
3.3 Conditions for Conducting the Examination.....	5
3.4 Test Design.....	5
3.5 Additional Tests.....	5
4. ASSESSMENT OF DISTINCTNESS, UNIFORMITY AND STABILITY.....	6
4.1 Distinctness.....	6
4.2 Uniformity.....	7
4.3 Stability.....	7
5. GROUPING OF VARIETIES AND ORGANIZATION OF THE GROWING TRIAL.....	8
6. INTRODUCTION TO THE TABLE OF CHARACTERISTICS.....	9
6.1 Categories of Characteristics.....	9
6.2 States of Expression and Corresponding Notes.....	9
6.3 Types of Expression.....	9
6.4 Example Varieties.....	9
6.5 Legend.....	10
7. TABLE OF CHARACTERISTICS/TABLEAU DES CARACTÈRES/MERKMALSTABELLE/TABLA DE CARACTERES.....	11
8. EXPLANATIONS ON THE TABLE OF CHARACTERISTICS.....	10
8.1 Explanations for individual characteristics.....	17
9. LITERATURE.....	19
10. TECHNICAL QUESTIONNAIRE.....	20

1. Subject of these Test Guidelines

These Test Guidelines apply to all varieties of *Phleum bertolonii* DC. and *Phleum pratense* L..

2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of seeds.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

500 grams

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

3. Method of Examination

3.1 *Number of Growing Cycles*

3.1.1 The minimum duration of tests should normally be two independent growing cycles.

3.1.2 The two independent growing cycles should be in the form of two separate plantings.

3.2 *Testing Place*

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

3.3 *Conditions for Conducting the Examination*

3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a number in the Table of Characteristics. The stages of development denoted by each number are described in Chapter 8.

3.3.3 The recommended type of plot in which to observe the characteristic is indicated by the following key in the Table of Characteristics:

A: spaced plants
B: row plots

3.4 *Test Design*

- 3.4.1 Spaced plants: Each test should be designed to result in at least 60 plants, which should be divided between at least 3 replicates.
- 3.4.2 Row plots: Each test should be designed to result in at least 3000 plants, which should be divided between at least 2 replicates.
- 3.4.3 The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.

3.5 *Additional Tests*

Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 *Distinctness*

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.1.4 Number of Plants or Parts of Plants to be Examined

Unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 60 plants or parts of plants taken from each of 60 plants and any other observations made on all plants in the test, disregarding any off-type plants.

In the case of observations of parts taken from single plants, the number of parts to be taken from each of the plants should be 1.

4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the Table of Characteristics (see document TGP/9 "Examining Distinctness", Section 4 "Observation of characteristics"):

MG: single measurement of a group of plants or parts of plants

MS: measurement of a number of individual plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

“Visual” observation (V) is an observation made on the basis of the expert’s judgment. For the purposes of this document, “visual” observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M) is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, “G” provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

4.2 *Uniformity*

4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:

4.2.2 These Test Guidelines have been developed for the examination of cross-pollinated varieties. For varieties with other types of propagation, the recommendations in the General Introduction and document TGP/13 "Guidance for new types and species" Section 4.5 "Testing Uniformity" should be followed.

4.2.3 The assessment of uniformity should be according to the recommendations for cross-pollinated varieties in the General Introduction.

4.3 *Stability*

4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.

4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.

5. Grouping of Varieties and Organization of the Growing Trial

5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.

5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.

5.3 The following have been agreed as useful grouping characteristics:

- (a) Plant: time of inflorescence emergence (characteristic 9)
- (b) Stem: length of longest stem (inflorescence included) (characteristic 15)

5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 “Examining Distinctness”.

6. Introduction to the Table of Characteristics

6.1 *Categories of Characteristics*

6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

6.2 *States of Expression and Corresponding Notes*

6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.

6.2.2 In the case of qualitative and pseudo-qualitative characteristics (see Chapter 6.3), all relevant states of expression are presented in the characteristic. However, in the case of quantitative characteristics with 5 or more states, an abbreviated scale may be used to minimize the size of the Table of Characteristics. For example, in the case of a quantitative characteristic with 9 states, the presentation of states of expression in the Test Guidelines may be abbreviated as follows:

<i>State</i>	<i>Note</i>
small	3
medium	5
large	7

However, it should be noted that all of the following 9 states of expression exist to describe varieties and should be used as appropriate:

<i>State</i>	<i>Note</i>
very small	1
very small to small	2
small	3
small to medium	4
medium	5
medium to large	6
large	7
large to very large	8
very large	9

6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 “Development of Test Guidelines”.

6.3 *Types of Expression*

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

6.4 Example Varieties

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

6.5 Legend

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1	2	3	4	5	6	7	
		Name of characteristics in English	Nom du caractère en français	Name des Merkmals auf Deutsch	Nombre del carácter en español		
		states of expression	types d'expression	Ausprägungsstufen	tipos de expresión		

- 1 Characteristic number
- 2 (*) Asterisked characteristic – see Chapter 6.1.2
- 3 Type of expression
 - QL Qualitative characteristic – see Chapter 6.3
 - QN Quantitative characteristic – see Chapter 6.3
 - PQ Pseudo-qualitative characteristic – see Chapter 6.3
- 4 Method of observation (and type of plot, if applicable)
 - MG, MS, VG, VS – see Chapter 4.1.5
- 5 (+) See Explanations on the Table of Characteristics in Chapter 8.1
- 6 Not applicable
- 7 Growth stage key See Explanations on the Table of Characteristics in Chapter 8.2

A = spaced plants

B = row plots

7. Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1.	QN VG B		20-29			
	Leaf: intensity of green color without vernalization					
	light					3
	medium					5
	dark					7
2.	QN VG B VS A	(+)	20-29			
	Plant: growth habit without vernalization					
	erect					1
	semi-erect					3
	medium					5
	semi-prostrate					7
	prostrate					9
3.	QN MG B VG B		20-29			
	Plant: natural height without vernalization					
	very low					1
	low					3
	medium					5
	high					7
	very high					9
4.	QN VS A	(+)				
	Plant: speed of inflorescence emergence without vernalization					
	absent or very slow				Vähäsoyrinki (P.p.)	1
	slow					3
	medium					5
	fast				Rubato (P.p.)	7
	very fast					9

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
5.	QN	VS A	(+)	20-29			
	Plant: growth habit after vernalization						
	erect						1
	semi-erect					Phlewiola (P.p.)	3
	medium					Barmidi (P.p.), Teno (P.b.)	5
	semi-prostrate					Latima (P.b.)	7
	prostrate						9
6. (*)	QN	MS A VG B		20-29			
	Plant: natural height after vernalization						
	very low						1
	low						3
	medium					Barmidi (P.p.)	5
	high					Tiller (P.p.)	7
	very high						9
7.	QN	VG B		30-39			
	Leaf: intensity of green color after vernalization						
	light						3
	medium						5
	dark						7
8.	QN	VG B		30-39			
	Leaf: width						
	narrow					Teno (P.b.)	3
	medium					Dolina (P.p.)	5
	wide						7
9. (*)	QN	MS A	(+)				
	Plant: time of inflorescence emergence						
	very early					Tiller (P.p.)	1
	early					Phlewiola (P.p.), Teno (P.b.)	3
	medium					Vähäsöyrinki (P.p.)	5
	late						7
	very late					Aberystwyth S48 (P.p.)	9

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
10.	QN	MS A VG B	50-56			
	Plant: natural height in time of inflorescence emergence					
	very low					1
	low					3
	medium					5
	high					7
	very high					9
11.	QN	VS A	50			
	Plant: growth habit at inflorescence emergence					
	erect					1
	semi-erect					3
	medium					5
	semi-prostrate					7
	prostrate					9
12. (*)	QN	MS A	(+)	52-56		
	Flag leaf: length					
	very short				Teno (P.b.)	1
	short					3
	medium					5
	long				Erecta (P.p.)	7
	very long					9
13. (*)	QN	MS A	52-56			
	Flag leaf: width					
	very narrow					1
	narrow					3
	medium				Tiller (P.p.)	5
	wide				KIS Muri (P.p.)	7
	very wide					9

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
14.	QN	MS A				
	Flag leaf: length/width ratio					
	very low					1
	low					3
	medium					5
	high					7
	very high					9
15. (*)	QN	MS A	60-68			
	Stem: length of longest stem (inflorescence included)					
	very short					1
	short					3
	medium				Vähäsoyrinki (P.p.)	5
	long				Dolina (P.p.)	7
	very long					9
16.	QN	MS A	60-68			
	Stem: length of upper internode	Tige: longueur du dernier entrenoeud	Halm: Länge des obersten Internodiums	Tallo: longitud del entrenudo superior		
	very short					1
	short				Teno (P.b.)	3
	medium	moyen	mittel	medio	Aberystwyth S48 (P.p.)	5
	long					7
	very long					9
17.	QN	MS A	60-68			
	Inflorescence: length (when fully expanded)	Inflorescence: longueur (à la fin de l'élongation)	Blütenstand: Länge (wenn voll ausgebildet)	Inflorescencia: longitud (cuando está plenamente desarrollada)		
	very short					1
	short				Teno (P.b.)	3
	medium	moyenne	mittel	media	Phlewiola (P.p.)	5
	long					7
	very long					9

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
18.	QN	MG B/VG B	30-39			
	Plant: natural height in aftermath					
	very low					1
	low					3
	medium					5
	high					7
	very high					9
19.	QN	VG B				
	Plant: intensity of inflorescence emergence in aftermath					
	very weak					1
	weak					2
	medium					3
	strong					4
	very strong					5

8.1 *Explanations for individual characteristics*

Ad. 2: Plant: growth habit without vernalization

The growth habit should be assessed visually from the attitude of the leaves of the plant as a whole. The angle formed by the imaginary line through the region of greatest leaf density and the vertical should be used.

Ad. 4: Plant: speed of inflorescence emergence without vernalization

Plant: speed of inflorescence emergence in the year of sowing

1st observation: - when approximately 50% of the plants of the earliest heading variety have emerged
- Note 8 for plants with emerged inflorescences

2nd observation: - 1-2 weeks after first observation (weather dependent)
- Note 6 for plants with emerged inflorescences

3rd observation: - 1-2 weeks after second observation (weather dependent)
- Note 4 for plants with emerged inflorescences

Note 2 for those other plants which have not emerged in any one of the three observations.
From these figures the descriptive states are calculated.

Ad. 5: Plant: growth habit after vernalization

The growth habit should be assessed visually from the attitude of the leaves of the plant as a whole. The angle formed by the imaginary line through the region of greatest leaf density and the vertical should be used.

Ad. 9: Plant: time of inflorescence emergence

The date of inflorescence emergence of each single plant should be assessed. A single plant is considered to have headed when the tip of three inflorescences can be seen protruding from the flag leaf sheath. From the single plant data a mean date per plot and a mean date per variety is obtained.

Ad. 12: Flag leaf: length

Flag leaf: The first true leaf at the top of the stem which is visible at the time of emerging inflorescence and has a sheath enclosing the stem.

Note: In some cases a small bract-like leaf which has a very short sheath, ligule and blade is visible directly at the base of the inflorescence. This leaf is not visible at the time of inflorescence emergence but is seen later as the inflorescence emerges. It generally does not have a normal sheath clasping the stem. This bract-like leaf is not to be considered as a true flag leaf.

8.2 *Growth stages for grasses*

All characteristics should be recorded at the appropriate time for the plant concerned. Growth stages of grasses are indicated by decimal codes which are derived from the decimal code for the growth stages of cereals (Zadoks, et al., 1974). This decimal code is in close conformity with the BBCH-code (Meier, 1997).

Seedling growth (seedling: one shoot)

DC 10 First leaf through coleoptile

DC 15 Five leaves unfolded

DC 19 Nine or more leaves unfolded

Tillering

DC 20 Main shoot only (beginning of tillering)

DC 23 Main shoot and 3 tillers

DC 25 Main shoot and 5 tillers

DC 29 Main shoot and 9 or more tillers

Stem elongation

DC 30 Pseudo-stem erection (formed by sheaths of leaves)

DC 31 First node detectable (early stem extension across all stems)

DC 35 Fifth node detectable (50 % extension across all stems)

DC 39 Flag leaf ligula/collar just visible (pre-boot stage)

Booting

DC 41 Flag leaf sheath extending (little enlargement of the inflorescence, early boot-stage)

DC 45 Boots swollen (late-boot stage)

DC 47 First leaf sheath opening

DC 49 first awns visible (in awned forms only)

Inflorescence emergence (mostly non-synchronous)

DC 50 First spikelet of inflorescence just visible

DC 52 25 % of the inflorescence emerged (across all stems)

DC 54 50 % of the inflorescence emerged (across all stems)

DC 56 75 % of the inflorescence emerged (across all stems)

DC 58 Emergence of inflorescence completed

Anthesis (mostly non-synchronous)

DC 60 Beginning of anthesis

DC 64 Anthesis half-way

DC 68 Anthesis complete

9. Literature

10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
		Application date: (not to be filled in by the applicant)
TECHNICAL QUESTIONNAIRE to be completed in connection with an application for plant breeders' rights		
1. Subject of the Technical Questionnaire		
1.1.1	Botanical name	<input type="text" value="Phleum bertolonii DC."/> []
1.1.2	Common name	<input type="text" value="Diploid Timothy, Small Timothy, Smaller Cat's-tail, Timothy, Turf Timothy"/>
1.2.1	Botanical name	<input type="text" value="Phleum pratense L."/> []
1.2.2	Common name	<input type="text" value="Meadow cat's-tail, Timothy"/>
2. Applicant		
	Name	<input type="text"/>
	Address	<input type="text"/>
	Telephone No.	<input type="text"/>
	Fax No.	<input type="text"/>
	E-mail address	<input type="text"/>
	Breeder (if different from applicant)	<input type="text"/>
3. Proposed denomination and breeder's reference		
	Proposed denomination (if available)	<input type="text"/>
	Breeder's reference	<input type="text"/>

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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#4. Information on the breeding scheme and propagation of the variety

4.1 Breeding scheme

Variety resulting from:

--

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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4.2	Method of propagating the variety	
4.2.1	Other (Please provide details)	[]
	<input type="text"/>	

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).

Characteristics	Example Varieties	Note
5.1 Plant: time of inflorescence emergence (9)		
very early	Tiller (P.p.)	1 []
early	Phlewiola (P.p.), Teno (P.b.)	3 []
medium	Vähäsoyrinki (P.p.)	5 []
late		7 []
very late	Aberystwyth S48 (P.p.)	9 []
5.2 Flag leaf: length (12)		
very short	Teno (P.b.)	1 []
short		3 []
medium		5 []
long	Erecta (P.p.)	7 []
very long		9 []
5.3 Flag leaf: width (13)		
very narrow		1 []
narrow		3 []
medium	Tiller (P.p.)	5 []
wide	KIS Muri (P.p.)	7 []
very wide		9 []
5.4 Stem: length of longest stem (inflorescence included) (15)		
very short		1 []
short		3 []
medium	Vähäsoyrinki (P.p.)	5 []
long	Dolina (P.p.)	7 []
very long		9 []

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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6. Similar varieties and differences from these varieties

Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.

Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the similar variety(ies)	Describe the expression of the characteristic(s) for your candidate variety
<i>Example</i>			
Comments:			

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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#7. Additional information which may help in the examination of the variety

7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?

Yes [] No []

(If yes, please provide details)

7.2 Are there any special conditions for growing the variety or conducting the examination?

Yes [] No []

(If yes, please provide details)

7.3 Other information

7.3.1 Ploidie
diploid []
hexaploid []

7.3.2 Resistance to pests and diseases
.....

7.3.2 Other
.....

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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8. Authorization for release

(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes [] No []

(b) Has such authorization been obtained?

Yes [] No []

If the answer to (b) is yes, please attach a copy of the authorization.

9. Information on plant material to be examined or submitted for examination

9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.

9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:

(a) Microorganisms (e.g. virus, bacteria, phytoplasma)	Yes []	No []
(b) Chemical treatment (e.g. growth retardant, pesticide)	Yes []	No []
(c) Tissue culture	Yes []	No []
(d) Other factors	Yes []	No []

Please provide details for where you have indicated "yes".

.....

10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:

Applicant's name

Signature Date

[End of document]